

Chemical and Functional Properties of Protein Isolate from Cowpea (*Vigna unguiculata*)

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Abstract— Cowpea (*Vigna unguiculata*) is potentially used as food ingredient since it has high protein content around 25%. This study was focused on the observation of the chemical and functional properties of cowpea protein isolate (CPI) compared to the chemical and functional properties of soy protein isolate (SPI) commercial with three repetitions in each parameter, then the data were analyzed descriptively. The results showed that CPI has the chemical properties of the moisture content, ash content, and carbohydrate content, which value is smaller than the value of SPI respectively 7.97%, 1.75%, 1.21%. CPI has protein and fat content which is higher, compared to SPI with consecutive values 88.06% and 1.05%. CPI contains more 7S globulin fraction compared with 11S and inversely related to SPI. CPI has functional properties including maximum solubility at pH 8, smaller foam capacity and higher foam stability than the value of SPI successive 68 ml/g and 8%, lower OHC and WHC than SPI at successive 84.89% and 136.61%, lower emulsion capacity and higher emulsion stability than the value of SPI with consecutive 2.41 m²/g and 78.15 hours, lower than the gelation of SPI with a value of 4 gf. CPI has a major fraction of protein bands with molecular weight of 59.11 kDa and 54.22 kDa, while the fraction of SPI has major protein bands with molecular weight of 50.66 kDa and 57.02 kDa.

Keywords— Cowpea Protein Isolate, Chemical And Functional Properties, 7s And 11s Globulin..

I. INTRODUCTION

Cowpea (*Vigna unguiculata*) has been widely known in Indonesia. Susilo and Imelda (2013) state that Cowpea production has reached 1,3 million tons. Every 100 gram-Cowpea material has nutrient content of 22.9 grams of protein, 1.1 grams of fat, and 61.6 grams of carbohydrates. This highly nutritional value makes cowpea deserved to be considered as a source of vegetable protein food to meet nutritional needs (Purwanti, 2010).

Protein isolate is the result of food protein isolation using the method of isolation of specific protein to produce the product with high protein concentration and pure. Chemical and functional properties such as solubility, 7S and 11S globulins, water holding capacity (WHC), oil holding capacity (OHC), emulsion properties, foam properties, gelation properties of protein isolates can improve the quality and organoleptic properties of food products. When the food industry is growing, the need for protein isolates will also increase.

Protein isolates are generally made from soybeans which are mostly imported from abroad. Data from the Central Statistics Agency (BPS) of Indonesia in 2013 showed a

fairly high value for imported raw materials of protein isolate was 1.192.173 tons. Though, Indonesia has the potential of local legumes protein source that can be used as raw material for the manufacture of protein isolates. The study of protein isolates from local materials has been developed but the results are less satisfactory. Mung bean protein isolates and fermented cowpea protein isolates only has protein content of 76.56% (Triyono, 2010) and 82.59% (Suproborini, 2011). Cowpea production is high enough to be used as an alternative raw material for making protein isolates. The technique of making cowpea protein isolates has been developed, but so far unknown chemical and functional properties of cowpea protein isolate as a scientific basis for further utilization for food ingredients.

II. MATERIALS AND METHODS

A. Materials

The basic material used for this research is cowpea derived from Malang, East Java and commercial soy protein isolate. Chemical materials used specification pro analysis Merck brands (Germany) covering 0.1N NaOH, 1N HCl, 0.5N HCl, 70% acetone, 0.1M phosphate buffer pH 7, 0.05M phosphate buffer pH 7, SDS 0.1%, Tris-HCl buffer

0.03 M pH 8, Lowry reagent, Follin, Temed, ammonium persulfate 10%, 30% polyacrylamide stock, 50% methanol, CBB R-250 0.05%, and glacial acetic acid 10 %.

B. Preparation of Cowpea Protein Isolate

The Preparation of CPI begins with cowpea weighing 100 grams and soaked in water for \pm 5 hours. Peeled epidermis and cuticle-free cowpea crushed using a blender and add distilled water ratio of 6:2 (water: materials). Thus, the obtained suspension was filtered in order to produce cowpea milk. Cowpea milk was added 0,1N NaOH in the ratio 5:1 (NaOH: suspension) and incubated at 55 °C for 30 minutes. Then, the milk was separated by using a centrifuge at 2000 rpm for 10 minutes. The supernatant was conditioned at pH 5 by the addition of 1N HCl and separated by using a centrifuge again at 2000 rpm for 10 minutes. While, the sediment is wet protein isolate purified using 70% acetone. After stirring with a stirrer was performed for 20 minutes and separation using a centrifuge at 2000 rpm for 10 minutes. The precipitate was dried using a vacuum oven at a temperature of 50 °C for 8 hours. CPI dried crushed and sieved using a 80 mesh sieve. After that, the cow pea protein isolate analyzed the chemical and functional properties compared to the properties of commercial soy protein isolates (SPI).

C. Chemical and Functional Properties Analysis

Chemical properties measured were moisture content (AOAC, 2005), ash content (AOAC, 2005), fat content (Soxhlet method, Sudarmadji, et al., 1997), protein content (Micro Kjedadhl, Sudarmadji, et al., 1997), carbohydrate content (By difference), the ratio of 7S and 11S globulins (Thanh and Shibasaki, 1976) and the composition of protein isolate fractions (Bollag et al., 1996). Functional properties of protein solubility was observed at various pH (Zayas, 1997), foam capacity and foam stability (Zayas, 1997), oil holding capacity (OHC) (Zayas, 1997), water holding capacity (WHC) (Zayas, 1997), emulsion capacity and stability (Parkington et al., 2000) and gelation (Dias et al., 2011).

D. Data Analysis

The data of chemical and functional properties of CPI were averaged over each parameter and analyzed descriptively. To interpret easily, the data were plotted in the table, histogram, or a picture.

III. RESULTS AND DISCUSSION

A. Chemical Properties of Cowpea Protein Isolate

The data of CPI and SPI chemical properties can be seen in Table 1.

TABLE I
CHEMICAL PROPERTIES OF CPI AND SPI

Parameters	CPI (%)	SPI (%)
Water	7.93 \pm 0.33	10.52 \pm 0.06
Ash	1.75 \pm 0.30	1.94 \pm 0.23
Fat	1.05 \pm 0.16	0.41 \pm 0.32
Protein	88.06 \pm 0.9	85.77 \pm 0.32
Carbohydrate	1.21 \pm 0.96	1.36 \pm 0.75

Table 1 shows that the CPI moisture content of 7.93 \pm 0.33 % while the moisture content SPI of 10.52 \pm 0.06%. High levels of moisture SPI was influenced by storage. SPI is only stored in plastic without any silica gel that absorbs moisture surrounding air. In accordance with the Triyono (2010), stated that the moisture content of the product depends on the packaging and storage time. Poor packaging could cause damage to the product and shelf life of the product would not be long. CPI moisture content in this study was not so different from isolates protein moisture content of green beans is equal to 7.39% (Triyono, 2010).

CPI ash content is 1.75 \pm 0.30% while the ash content SPI is 1.94 \pm 0.23% (Table 1). CPI ash content values is lower than the SPI because the ash content of cowpea is also lower than that of soybean ash content of 1.97% and 2.14% (Danuwarsa, 2010). CPI ash content in this study is lower than the ash content of jack bean protein isolate that is equal to 2.66% (Subagio et al., 2003).

Table 1 shows that the CPI fat content is 1.05 \pm 0.16 % while the SPI fat content is 0.41 \pm 0.32%. High fat content in CPI allegedly due to fat solvents used in the manufacturing process of CPI is acetone what is not so maximum dissolves the fat so that the fat content of CPI is higher. Also according to Iskandar (2003), SPI is made from fat-free soy flour to produce a protein isolate with a lower fat content.

CPI protein content is 88.06 \pm 0.96% and SPI protein content is 85.77 \pm 0.32% (Table 1). CPI has higher protein content than the SPI. CPI protein content of these results is far different from mung bean protein isolates and fermented cowpea protein isolates are equal to 76.56% (Triyono, 2010) and 82.59% (Suproborini, 2011). In the fermented cowpea protein isolates, proteins already hydrolyzed since the fermentation process in order the protein content decreased.

CPI carbohydrate content is 1.21 \pm 0.96% and SPI carbohydrate content is 1.36 \pm 0.75% (Table 1). CPI has lower carbohydrate content than the SPI. The results of this study also showed that the carbohydrate content of CPI is lower carbohydrate content than the jack bean protein isolate. Subagio et al. (2003) reported that the carbohydrate content of jack bean protein isolates is 21.48%.

The ratio of 7S and 11S globulins contributes to the robustness and elasticity of the gel formed in food. The results of the analysis of 7S and 11S globulin ratio CPI and SPI can be seen in Table 2 below.

TABEL II
RATIO OF 7S AND 11S GLOBULINS CPI AND SPI

Sampel	Globulin levels sample type (%)	
	7S Globulin	11S Globulin
CPI	0.03 \pm 0.03	0.02 \pm 0.00
SPI	0.05 \pm 0.01	0.25 \pm 0.07

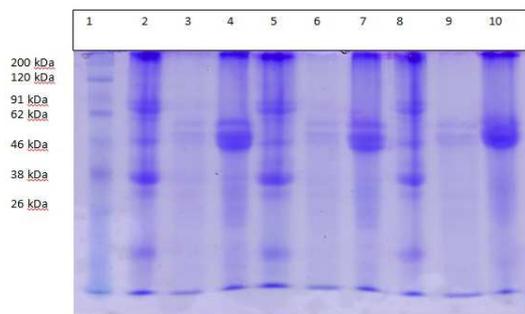


Fig. 1 Results of SDS-PAGE electrophoresis (1: marker; 2, 5, 8: SPI; 3, 6, 9: CPI dry; 4, 7, 10: CPI wet)

Figure 1 shows that the SPI has 6 bands with molecular weight of protein fraction in a row is 79.23; 80.45; 60.57; 52.03; 33.88 and 22.43 kDa. Protein fraction with a molecular weight of 80.45 and 33.88 kDa is the major protein fraction. CPI dried protein fraction has 5 bands with successive molecular weight of 65.67; 59.11; 54.22; 37.57 and 32.11 kDa. Major protein fraction is the fraction of proteins with molecular weight of 59.11 and 54.22 kDa. Wet CPI has 8 bands of protein fractions by successive molecular weight of 65.53; 57.02; 50.66; 37.06; 31.56; 27.10; 24.94 and 22.86 kDa. Protein fraction with a molecular weight of 50.66 and 57.02 kDa is the major protein fraction. In accordance with research Horax et al. (2004) which states that the cowpea protein has a molecular weight of major proteins ranged from 40 to 66.2 kDa.

B. Functional Properties of Cowpea Protein Isolate

The Functional properties analyzed include protein solubility at various pH, foam capacity and foam stability, oil holding capacity (OHC), water holding capacity (WHC), emulsion capacity and emulsion stability, and gelation.

C. The Solubility of Proteins at Various pH

Figure 2 below shows that the highest protein solubility of CPI and SPI at various pH is at 8, which means that the pH of the protein can be dissolved as much as on pH above or below the isoelectric point, the protein is changed so that the charge affinity between the protein molecules decreased. This causes the protein molecules readily biodegradable and will increase the solubility of the protein. Ragap et al. (2004) reported that cowpea protein isolate have highly soluble at alkaline pH.

According to Kinsella (1985) in Aluko and Yada (1995), at pH above and below the isoelectric point, the protein will change the charge causes a decreased affinity between protein molecules, so the molecules are more easily broken down.

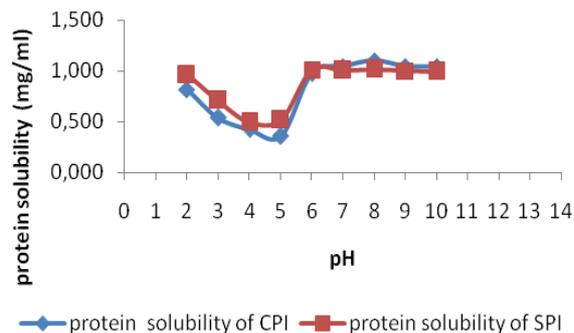


Fig. 2 CPI and SPI protein solubility at various pH

D. Foam Capacity and Foam Stability

Figure 3 below shows that the foam capacity of CPI is 68 ± 4.00 ml/g, while the foam capacity of SPI by 136 ± 6.93 ml/g. The low foam capacity of CPI influenced by the composition fraction of protein isolate. SPI has a more complex composition fraction compared to CPI. It is thought bringing the fact to the foam capacity. In accordance with Alleoni and Antunes (2004) which states that one of the factors affecting the foam capacity that the protein composition. The protein composition of CPI and SPI is different that it causes foam capacity different, too.

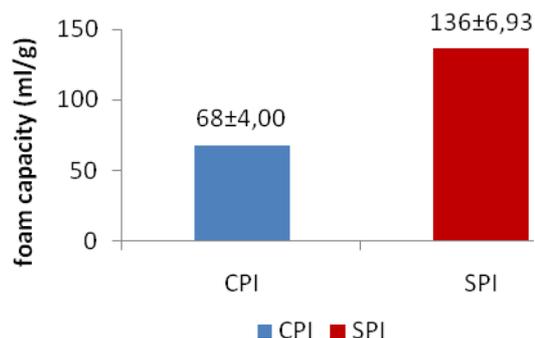


Fig. 3 CPI and SPI foam capacity

Figure 4 below shows that the foam stability of CPI is $8 \pm 0.00\%$ and the foam stability of SPI is $6 \pm 2.00\%$. The high foam stability of CPI is influenced by the protein content. CPI has a higher protein content than the SPI so the foam stability more high. In accordance with Aluko and Yada (1995), which state that the foam stability is influenced by the protein content.

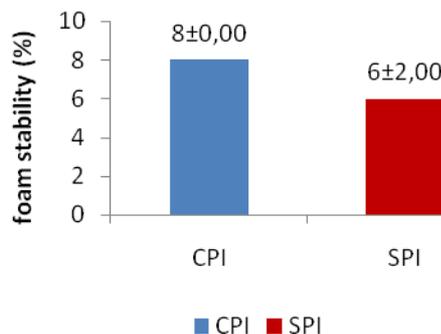


Fig. 4 CPI and SPI foam stability

E. Oil Holding Capacity (OHC)

Figure 5 below shows that the oil absorption of CPI is $84.89 \pm 1.36\%$, while the oil absorption of SPI is $121.07 \pm 14.40\%$. The low oil absorption of CPI affected particle size. SPI has a particle size smaller than CPI. According to Iskandar (2003), the nature off at absorption is influenced by the size of the protein particles. The particle size and finer texture and uniform lead are more easily absorbed protein isolates and oil binding.

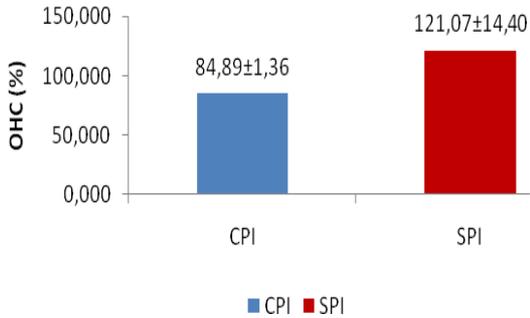


Fig 5. CPI and SPI oil holding capacity (OHC)

F. Water Holding Capacity (WHC)

Water Holding Capacity is as shown in Figure 6 below.

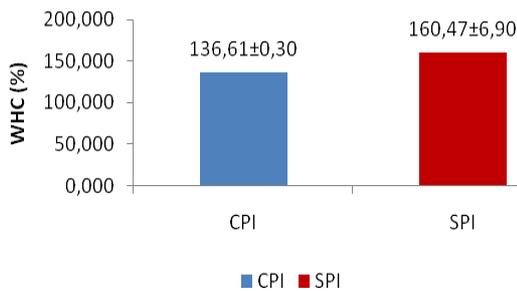


Fig. 6 CPI and SPI water holding capacity (WHC)

This figure shows that the water absorption of CPI is $136.61 \pm 0.30\%$ while the water absorption of SPI is $160.47 \pm 6.90\%$. The low water absorption of CPI associated with 11S globulin fraction. 11S globulin fractions of CPI have less than the SPI so the water absorption of CPI is lower. This is consistent with Yowono et al. (2004) which states that the 11S globulin may improve the ability to bind water (WHC). Horax et al., (2004) reported that cowpea protein had lower hydrophobicity so the ability to bind water is lower.

G. Emulsion Capacity and Emulsion Stability

Figure 7 below shows that the emulsion capacity of CPI is 2.41 ± 0.16 m²/g and emulsion capacity of SPI of 3.74 ± 2.06 m²/g. Low emulsion capacity on CPI allegedly due to the composition of the different fractions of CPI and SPI. SPI has a composition of protein fractions that are more complex than CPI. According Alleoni and Antunes (2004), one of the factors influencing the emulsion capacity is composition of proteins. SPI has a composition more complex protein fraction so that emulsion capacity gets higher. Ragap et al. (2004) reported that emulsion capacity influence the composition of proteins.

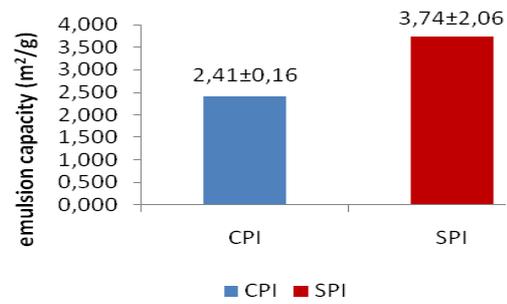


Fig. 7 CPI and SPI emulsion capacity

Figure 8 below shows that the emulsion stability of CPI and SPI, respectively for 78.15 ± 2.60 hours and 72.74 ± 4.15 hours. Emulsion stability is related to the 7S and 11S globulins contained in CPI and SPI. CPI 7S globulin which has led to higher emulsion stability is also high. This is consistent with Aoki, Taneyama, and Inami (1980) in Zayas (1997) who state that the 7S protein fractions have better emulsion stability compared to 11S protein fraction.

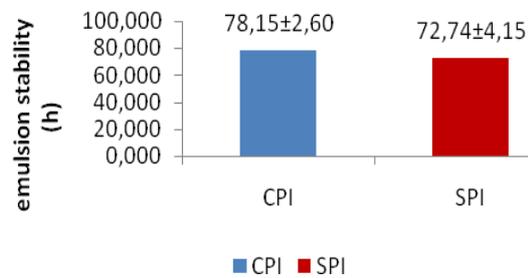


Fig. 8 CPI and SPI emulsion stability

H. Gelation

Figure 9 (1) below shows the gelation of CPI is 4.00 ± 0.00 gf and the gelation of SPI is 5.00 ± 0.00 gf. Figure 9 (2) shows the gelation of CPI and SPI after being stored in the refrigerator for 48 hours in a row amounted to 5.53 ± 0.90 and 7.87 ± 0.58 gf. This indicates that the gel formed from CPI softer than the SPI because the larger value indicated that the gel formed will be the hardest. Cooling treatment also increases the hardness of gel formed. Gelation properties of proteins often associated with the presence of 7S and 11S proteins. CPI 7S globulin protein has greater than SPI, while SPI has 11S globulin proteins larger than CPI. This is consistent with Corredig (2006) which states that the gel obtained from isolation glicinin (11S) gives the character a harder gel than gels obtained from β -conglisinin (7S), and the network structure is formed to have a difference between the two, depending of protein composition.

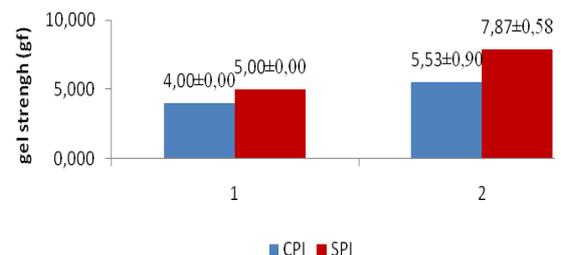


Fig. 9 (1) Gelation of CPI and SPI; (2) Gelation of CPI and SPI after being stored in the refrigerator about 48 hours

IV. CONCLUSIONS

CPI has chemical properties including moisture content, ash content, carbohydrate content, protein content and fat content with value respectively 7.97%, 1.75%, 1.21%, 88.06% and 1.05%. More CPI containing 7S globulin fraction compared with 11S. CPI has functional properties include solubility in a wide range of pH that dissolves at pH 8, foam capacity and foam stability, respectively for 68 ml/g and 8%, OHC and WHC with values respectively 84.89% and 136.61%, emulsion capacity and emulsion stability in a row by 2,41 m²/g and 78,15 hours, and gelation of 4 gf. CPI has a major fraction of protein band with molecular weight of 59.11 kDa and 54.22 kDa.

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