







one and five weeks incubated peat and 5 weeks incubated tapioca flour significant results were observed.

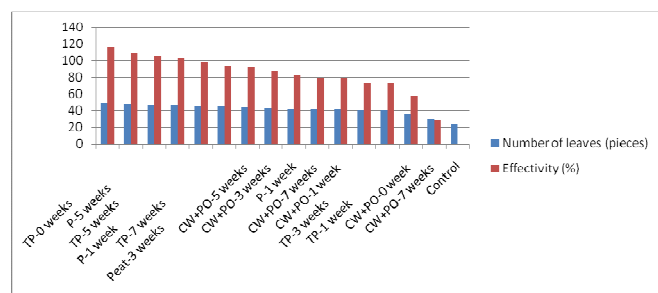


Fig. 4. Number of leaves on rhizobacterial introduced soybean and their effectivity

The days required by plants to reach flowering was significantly advanced by seven days faster in rhizobacterial introduced soybean compare than control, such as three, five and seven weeks incubated coconut water formula, three, five and seven weeks incubated tapioca flour formula, and one, three, and seven weeks incubated peat formula (Table 2).

TABLE II  
BEGIN OF FLOWERING PHASE OF RHIZOBACTERIAL INTRODUCED SOYBEAN AND THEIR EFFECTIVITY

Formulation- Incubation periode (weeks)	Generative phase (DAP)	Effectivity (%)
Coconut water+palm oil 0 week	47,67	-3,63
Control	46,00	0,00
Tapioca flour-1 weeks	44,67	2,89
Peat-5 weeks	44,00	4,35
Coconut water+palm oil-1 weeks	43,33	5,80
Peat-0 week	43,33	5,80
Peat-1 week	41,00	10,87
Peat-7 weeks	40,33	12,33
Coconut water+palm oil - 7 weeks	40,00	13,04
Tapioca flour-0 week	40,00	13,04
Coconut water+palm oil - 5 weeks	40,00	13,04
Tapioca flour -5 weeks	39,33	14,50
Tapioca flour -3 weeks	39,33	14,50
Peat-3 weeks	39,33	14,50
Tapioca flour -7 weeks	39,33	14,50
Coconut water+palm oil 3 week	39,00	15,22
SD = 9,69 %		

Under greenhouse conditions the soybean yield was significantly increased by fast all the formulations and incubation periode compared with the nontreated control. In case of increase in soybean yield, there was significant difference among the different formulations and incubation

periodes. All of them were significantly higher than control (Figure 5).

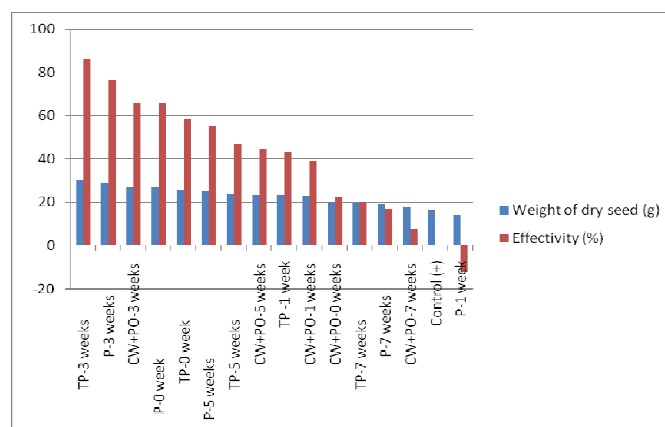


Fig. 5. Soybean yield after application of rhizobacterial formula and different incubation periode and their effectivity.

### B. Discussion

We The ability of PGPR in growth promotion and resistance induction in various crops is well documented ([17], [18], [19], [20]). The results reported here corroborate earlier studies and indicate a future possibility that PGPR formulations can be used to promote growth of crop plants. Treatments with rhizobacterial formulations significantly enhanced the growth of soybean plants.

Peat formulations have also promoted growth in different plants like cucumber, watermelon, squash, ornamentals, vegetables, pepper, tobacco and tree species like loblolly pine, and lodge pine Douglas fir. ([21], [22], [23], [24], [25], [26], [27]). However, this is the first report that demonstrates the efficiency of tapioca flour and coconut water formulations promoted growth in soybean plant.

Our results suggest that PGPR formulation can be used practically in production of soybean. The practical applications of these formulations were supported by the magnitude of growth promotion recorded by these treatments which was highly significant in comparison with the nontreated control. The most important result was the considerable increase in yield of soybean. Another important result was the advancement of flowering faster by 7–8 days than non treated control.

Earlier studies on PGPR have also reported that rhizobacteria are potential growth enhancers in different crops like potato, pearl millet, and sorghum ([28], [29], [30]). In the present study, all the formulations tested showed their efficacy in enhancing germination of soybean. The treated plants showed advanced emergence of seedlings in comparison to the control. In general, all the formulations showed a significant enhancement of growth and reproductive parameters such as height, number of leaf, flowering stage, and yield of soybean under greenhouse conditions

#### IV. CONCLUSIONS

All the formulations showed a significant enhancement of growth and reproductive parameters such as height, number of leaf, flowering stage, and yield of soybean under greenhouse conditions.

#### ACKNOWLEDGMENT

This research was funded by DP2M-DIKTI DEPDIKNAS (Hibah Kompetensi grant), Contract Nr. 134/UN16/PL/2011, April 21, 2012. We would like to sincerely thanks to Director of DP2M-DIKTI.

#### REFERENCES

- [1] M. Stefan, S. Dunca, Z. Olteanu, L. Oprica, E. Ungureanu, L. Hritcu, M. Mihasan & D. Cojocaru. Soybean (*Glycine max* [L] Merr.) inoculation with *Bacillus pumilus* RS3 promotes plant growth and increases seed protein yield: relevance for environmentally-friendly agricultural applications. Carpathian Journal of Earth and Environmental Sciences, Vol. 5, No. 1, p. 131 – 138, April 2010.
- [2] M. Stefan, M. Mihasan, L. Raus, D. Topa, S. Dunca. Agriculture applications of some rhizobacterial strains isolated from Moldavian Plaine Cambic-Chernozemic soils. Lucrări Științifice –Vol. 51:191-196, seria Agronomie, 2009.
- [3] B. R. Glick. The enhancement of plant growth by free living bacteria. Can J Microbiol 41: 109-114, 1995.
- [4] B. R. Glick, Z. Cheng, J. Czarny and J. Duan, Promotion of plant growth by ACC deaminase containing soil bacteria. Eur. J. Plant Pathol., 119: 329-339, 2007b.
- [5] B. R. Glick, B. Todorovic, J. Czarny, Z. Cheng, J. Duan and B. McConkey, 2007a. Promotion of plant growth by bacterial ACC deaminase. Crit. Rev. Plant Sci., 26: 227-242, 2007a.
- [6] H. Antoun, and J. W. Kloepper, Plant Growth Promoting Rhizobacteria. Encyclopedia of Genetics. London: Academic Press, 2001.
- [7] R. M. N. Kucey, H. H. Janzen, and M. E. Leggett, Microbiologically Mediated Increases in Plant-Available Phosphorus. Advances in Agronomy, 42, 199-228, 1989.
- [8] Y. Yanti, T. Habazar, Z. Resti and D. Suhailita, Penapisan isolat rizobakteri dari perakaran tanaman kedelai yang sehat untuk pengendalian penyakit pustul bakteri (*Xanthomonas axonopodis* pv. *glycines*). J. HPT Tropika 13(1):24-34, 2013
- [9] U. Chakraborty, B. N. Chakraborty and A. P. Chakraborty, Induction of plant growth promotion in *Camellia sinensis* by *Bacillus megaterium* and its bioformulations. World Journal of Agricultural Sciences 8 (1): 104-112. ISSN 1817-3047, 2012.
- [10] S. Nakkeeran, K. Kavitha, G. Chandrasekar, P. Renukadevi, and W. G. D. Fernando, Induction of plant defence compounds by *Pseudomonas chlororaphis* PA23 and *Bacillus subtilis* BSCBE4 in controlling damping-off of hot pepper caused by *Pythium aphanidermatum*. Biocontrol Science and Technology; 16(4): 403-416, 2006
- [11] R. D. Lumsden, J. A. Lewis and D. R. Fravel, Formulation and delivery of bio-control agents for use against soil-borne plant pathogens. In: F.R. Hall Formulation and Delivery, Washington, DC, USA, American Chemical Society, pp: 166-182, 1995.
- [12] S. S. Ardakani, A. Heydari, N. Khorasani and R. Arjmandi, Development of new bioformulations of *Pseudomonas fluorescens* and evaluation of these products against damping-off of cotton seedlings. Journal of Plant Pathology 92 (1), 83-88, 2010.
- [13] D. C. Thompson, Evaluation of bacteria immunization: an alternative to pesticides for control of plant disease in greenhouse and field. In: Bay-Petersen J. (ed.). The Biological Control of Plant Disease, pp. 30-40. Fertilizer Technology Centre, Taiwan, 1996.
- [14] ISTA, Proceedings of the international Seed Testing Association, International Rules for Seed Testing. Seed Sci. Technol. 21, pp. 25–30. 1993.
- [15] A. A. Abdul Baki and J. D. Anderson, Vigor determination in soybean seed by multiple criteria. Crop Sci. 13, 630–633, 1973.
- [16] P. Vidhyasekaran, P. R. Rabindran, M. Muthamilan, K. Nayar, K. Rajappan, N. Subramanian, K. Vasumathi, Development of a powder formulation of *Pseudomonas fluorescens* for control of rice blast. Plant Pathology 46:291-297, 1997.
- [17] L. C. van Loon, P. A. H. M. Baker, and C. M. J. Pieterse, Systemic resistance induced by rhizosphere bacteria. Annu Rev Phytopathol 36:453-483, 1998.
- [18] E. A. Barka, A. Belarbi, C. Hachet, J. Nowak, J. Audran, Enhancement of in vitro growth and resistance to gray mold of *Vitis vinifera* co-cultured with plant growth-promoting rhizobacteria. FEMS Microbiol. Lett. 186, 91–95, 2000.
- [19] S. Burdman, E. Turkevitch, and Y. Okon, Recent advances in the use of plant growth promoting rhizobacteria (PGPR) in agriculture. In: Subba Rao, N.S., Dommergues, Y.R. (Eds.), Microbial Interaction in Agriculture and Forestry. Vol. 2., Science Publishers Inc, Enfield (NH), USA, pp. 229–249, 2000.
- [20] V. Ramamoorthy, R. Viswanathan, T. Raguchander, V. Prakasam, R. Samiyappan, Induction of systemic resistance by plant growth promoting rhizobacteria in crop plants against pests and diseases. Crop Protection 20:1-11, 2001.
- [21] M. S. Reddy, R. Rodriguez-Kabana, D. S. Kenney, C. M. Ryu, S. Zhang, Z. Yan, N. Martinez-Ochoa, J. W. Kloepper, Growth promotion and induced systemic resistance (ISR) mediated by a biological preparation. Phytopathology 89, S65, 1999.
- [22] D. S. Kenney, M. S. Reddy, J. W. Kloepper, Commercial potential of biological preparations for vegetable transplants. Phytopathology 89, S39, 1999.
- [23] J. W. Kloepper, R. Rodriguez-Kabana, D. S. Kenney, M. S. Reddy, N. Martinez-Ochoa, N. Kokalis-Burelle, K. Arthur, Development of an integrated biological approach to develop transplants suppressive to various plant diseases. Phytopathology 89, S40, 1999.
- [24] N. Martinez-Ochoa, N. Kokalis-Burelle, R. Rodriguez-Kabana, J. W. Kloepper, Use of organic amendments, botanical aromatics, and rhizobacteria to induce suppressiveness of tomato to the root-knot nematode, *Meloidogyne incognita*. Phytopathology 89, S49, 1999.
- [25] C. M. Ryu, M. S. Reddy, S. Zhang, J. F. Murphy, J.W. Kloepper, 1999. Plant growth promotion of tomato by a biological preparation (LS 213) and evaluation for protection against cucumber mosaic virus. Phytopathology 89, S87. ev and V. P. Veiko, Laser Assisted Microtechnology, 2nd ed., R. M. Osgood, Jr., Ed. Berlin, Germany: Springer-Verlag, 1998.
- [26] Z. Yan, M. S. Reddy, Q. Wang, R. Mei, J. W. Kloepper, Role of rhizobacteria in tomato early blight control. Phytopathology 89, S87, 1999.
- [27] S. Zhang, M. S. Reddy, C. M. Ryu, J. W. Kloepper, Relationship between in vitro and in vivo testing of PGPR for induced systemic resistance against tobacco blue mold. Phytopathology 89, S89, 1999.
- [28] G. Lazarovitz, and J. Nowak, Rhizobacteria for improvement of plant growth and establishment. Hort. Sci. 32, 92–96, 1997.
- [29] S. Umesh, S. M. Dharmesh, S. A. Shetty, M. Krishnappa, H. S. Shetty, Biocontrol of downy mildew disease of pear millet using *Pseudomonas fluorescens*. Crop Prot. 17, 387–392, 1999.
- [30] N. S. Raju, S. R. Niranjana, G. R. Janaradhana, H. S. Prakash, H. S. Shetty, S. B. Mathur, Improvement of seed quality and field emergence of *Fusarium moniliforme* infected sorghum seeds using biological agents. J. Sci. Food Agri. 79, 206–212, 1999.